

# Determination of Parent and Substituted Polycyclic Aromatic Hydrocarbons in High-Fat Salmon Using a Modified QuEChERS Extraction, Dispersive SPE and GC–MS

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**ABSTRACT:** A fast and easy modified QuEChERS (quick, easy, cheap, rugged and safe) extraction method has been developed and validated for determination of 33 parent and substituted polycyclic aromatic hydrocarbons (PAHs) in high-fat smoked salmon that greatly enhances analyte recovery compared to traditional QuEChERS procedures. Sample processing includes extraction of PAHs into a solution of ethyl acetate, acetone and isooctane followed by cleanup with dispersive SPE and analysis by GC–MS in SIM mode. Method performance was assessed in spike recovery experiments (500  $\mu\text{g/g}$  wet weight) in three commercially available smoked salmon with 3–11% fat. Recoveries of some 2-, 3- and 5-ring PAHs were improved 50–200% over traditional methods, while average recovery across all PAHs was improved 67%. Method precision was good with replicate extractions typically yielding relative standard deviations <10%, and detection limits were in the low ng/g range. With this method, a single analyst could extract and clean up  $\geq 60$  samples for PAH analysis in an 8 h work day.

**KEYWORDS:** PAHs, polycyclic aromatic hydrocarbons, QuEChERS, smoked salmon, fish, seafood, biomatrix, food safety

## INTRODUCTION

Polycyclic aromatic hydrocarbons (PAHs) and their substituted derivatives are widespread environmental contaminants that may originate from pyrogenic or pyrogenic sources. PAHs are present in smoke as combustion byproducts<sup>1–3</sup> and display a range of toxic, mutagenic and carcinogenic properties.<sup>4,5</sup> It has been demonstrated that PAH burdens are generally higher in smoked foods than in the corresponding non-smoked foods<sup>6–8</sup> and that PAH profiles in foods are combustion material specific.<sup>9</sup> Bioanalytical methods that allow for the rapid monitoring of these residues in lipid-rich foods are therefore important for monitoring human dietary exposure to PAHs.

Analytical methods have been developed for assessing PAH residue loads in many matrices with the distinguishing factor being the mode of sample preparation. Common preparation techniques include solid–liquid extraction, Soxhlet extraction, sonication assisted extraction or accelerated solvent extraction coupled to a sample cleanup procedure using solid-phase extraction or gel permeation chromatography.<sup>6–13</sup> However, these methods are often labor, time and solvent intensive, require advanced analytical expertise and lab equipment and rely on the use of chlorinated extraction solvents. To overcome these challenges, QuEChERS (quick, easy, cheap, effective, rugged and safe) based sample processing procedures have been investigated.

Traditionally, the QuEChERS method has been used for the rapid (<15 min/batch) determination of pesticide residues in fruits and vegetables where sample extraction involves addition of acetonitrile and subsequent liquid–liquid partitioning of residues through the addition of magnesium sulfate, sodium chloride and various pesticide specific pH buffering agents such as sodium acetate or sodium citrate. Sample cleanup is then achieved using various dispersive solid-phase extraction materials and magnesium sulfate to remove polar matrix components and water.<sup>14</sup> The streamlined nature of the QuEChERS procedure has led to

its implementation in the analysis of veterinary pharmaceuticals in animal tissues, mycotoxins in breakfast cereals and flours, phthalates in fruit jellies and oil dispersion surfactants used in the 2010 Deepwater Horizon oil spill.<sup>15–19</sup> Furthermore, several QuEChERS methods have been developed for the analysis of PAHs in seafood such as shrimp, scallops, mussel and finfish.<sup>20–25</sup>

Though several QuEChERS based PAH methods have been previously described, none have been validated in high-fat biomatrices (>3.5% fat) or with substituted PAHs. Application of traditional QuEChERS methods to PAH extraction from high-fat salmon led to poor recoveries, typically averaging only 61–68%. Most QuEChERS methods are validated using only 16 EPA priority pollutant PAHs in low-fat National Institute of Standards and Technology standard reference materials, such as mussel tissue. Given the unmet need for a robust PAH method in high-fat biomatrices, we sought to develop a fast, selective and sensitive analytical method that combined the QuEChERS high throughput attributes with an extended characterization of PAHs in high-fat biomatrices. Using two developed modified QuEChERS methods, three high-fat salmon were characterized for 33 PAHs. The fat profile ranged from 3 to 11% fat, the highest known values reported for a QuEChERS based PAH extraction method.

## MATERIALS AND METHODS

**Chemicals and Food Products.** A stock solution of 33 PAHs and substituted PAHs was prepared by combining 16 EPA priority pollutant PAHs, a custom PAH mix and individual PAHs and diluting to volume with isooctane (see Table 1). In addition to EPA priority pollutant PAH residues, selected PAHs included retene for use as a marker of certain

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**Table 1. Retention Times, Monitored Quantitation and Confirmation Ions, and Instrument Detection Limits (IDL) for 33 PAHs by GC–MS**

PAH	corresp chromatogram no.	DB5 $t_R$ (min)	target compd monitored SIM ions ( $m/z$ )		$r^2$	IDL <sup>a</sup> (pg/ $\mu$ L)
			quant	confirm		
perylene-D12	ISTD 1	25.72	264	260, 265		
naphthalene	1	8.77	128	127, 129	0.999	1
2-methylnaphthalene	2	10.30	142	141, 115	0.999	1
1-methylnaphthalene	3	10.52	142	141, 115	0.999	1
1,6-dimethylnaphthalene	4	11.97	156	141, 153	0.999	1
acenaphthylene	5	12.34	152	151, 150	0.999	1
1,2-dimethylnaphthalene	6	12.35	141	156, 115	0.999	1
acenaphthene	7	12.75	153	154, 152	0.999	1
fluorene	8	13.96	166	165, 167	0.999	1
dibenzothiophene	9	15.91	184	139, 185	0.999	1
phenanthrene	10	16.21	178	176, 179	0.999	5
anthracene	11	16.33	178	176, 179	0.996	5
2-methylphenanthrene	12	17.42	192	191, 165	0.998	5
2-methylanthracene	13	17.53	192	191, 165	0.992	5
1-methylphenanthrene	14	17.67	192	191, 165	0.999	5
9-methylanthracene	15	18.00	192	191, 165	0.999	5
3,6-dimethylphenanthrene	16	18.43	206	191, 205	0.999	1
fluoranthene	17	19.01	202	203, 200	0.999	1
2,3-dimethylanthracene	18	19.09	206	191, 205	0.996	5
pyrene	19	19.53	202	200, 203	0.999	1
9,10-dimethylanthracene	20	19.60	206	191, 205	0.998	1
retene	21	20.27	219	220, 234	0.999	1
1-methylpyrene	22	20.86	216	215, 217	0.999	1
benz[ <i>a</i> ]anthracene	23	22.37	228	226, 229	0.983	5
chrysene	24	22.45	228	226, 229	0.994	1
6-methylchrysene	25	23.49	242	241, 226	0.999	1
benzo[ <i>b</i> ]fluoranthene	26	24.78	252	253, 250	0.994	1
benzo[ <i>k</i> ]fluoranthene	27	24.85	252	253, 250	0.992	1
benzo[ <i>e</i> ]pyrene	28	25.44	252	250, 253	0.999	1
benzo[ <i>a</i> ]pyrene	29	25.57	252	253, 250	0.996	1
indeno[1,2,3- <i>cd</i> ]pyrene-D12	ISTD 2	28.78	288	284, 289		
indeno[1,2,3- <i>cd</i> ]pyrene	30	28.86	276	277, 274	0.997	1
dibenz[ <i>a,h</i> ]anthracene	31	28.95	278	279, 276	0.998	5
benzo[ <i>g,h,i</i> ]perylene	32	29.63	276	277, 274	0.999	1
dibenz[ <i>a,l</i> ]pyrene	33	33.91	302	300, 303	0.998	1

<sup>a</sup> IDL assigned when PAH lowest abundant confirmation ion  $S/N > 3$  for standards prepared in isoctane.

types of wood combustion material,<sup>1</sup> substituted PAHs due to high levels reported in smoked fish<sup>7</sup> and dibenzo[*a,l*]pyrene because of its recent identification in NIST SRMs associated with urban combustion pollution.<sup>26</sup> Furthermore, exposure to these compounds has been associated with an increased risk for developing adverse health effects.<sup>26,27</sup> All premade mixtures and individual PAHs were purchased from AccuStandard Inc. (New Haven, CT) and were guaranteed to be greater than 97% pure. Working standards were prepared by dilution of the stock standard with isoctane and stored in the dark at 4 °C.

Perylene-D12 and indeno[1,2,3-*cd*]pyrene-D12 were purchased from Cambridge Isotope Laboratories, Inc. (Andover, MA) and used as internal standards for instrumental quantitation. High purity Optima acetonitrile, ethyl acetate, acetone, hexane and pesticide grade isoctane were purchased from Fisher Scientific (Pittsburgh, PA) and used throughout the study. Glacial acetic acid was from J.T. Baker (Phillipsburg, NJ), and high purity water was supplied by a Barnstead EASYpure UV compact

ultrapure water system (Dubuque, IA). Commercially available Sampli-Q QuEChERS AOAC (6 g of magnesium sulfate, 1.5 g of sodium acetate/package) and EN (4 g of magnesium sulfate, 1 g of sodium chloride, 1 g of sodium citrate, 0.5 g of sodium hydrogen citrate sesquihydrate/package) extraction salts and 2 mL of AOAC fatty sample dispersive SPE tubes (50 mg of PSA, 50 mg of C18EC and 150 mg of magnesium sulfate) were obtained from Agilent Technologies (Santa Clara, CA).

Three commercially available smoked salmon fillets (~113 g wet weight) were purchased from local grocery stores. Salmon fat content was determined and reported by the manufacturer in the product nutrition label. Salmon were homogenized via freeze fracture using a Robot Coupe Blixer 2 food processor (Ridgeland, MS) and liquid N<sub>2</sub>, transferred into amber glass screw-top jars and stored in the dark at -20 °C. All sample preparation equipment and machinery were washed with soap and water and rinsed with high purity water, acetone and hexane prior to use and between samples.

Table 2. Summary of Tested QuEChERS Extraction Conditions for Recovery of 33 PAHs from Smoked Salmon

extraction scheme	extraction solvent (vol)	extraction salt	
		type and composition (g)	amount used (g)
E1	1% (v/v) acetic acid in acetonitrile (5 mL)	AOAC <sup>a</sup> : MgSO <sub>4</sub> (6 g), NaC <sub>2</sub> H <sub>3</sub> O <sub>2</sub> (1.5 g)	2.5
E2	acetonitrile (2 mL)	EN <sup>b</sup> : MgSO <sub>4</sub> (4 g), NaCl (1 g), NaC <sub>6</sub> H <sub>7</sub> O <sub>7</sub> (1 g), Na <sub>2</sub> C <sub>6</sub> H <sub>8</sub> O <sub>8</sub> (0.5 g) <sup>c</sup>	1.3
E3	2:2:1 (v/v/v) acetone, ethyl acetate, isooctane (2 mL)	AOAC	1.3
E4	2:2:1 (v/v/v) acetone, ethyl acetate, isooctane (2 mL)	EN	1.3

<sup>a</sup> Commercially available extraction salt packets developed by the Association of Official Analytical Chemists (AOAC). <sup>b</sup> Commercially available extraction salt packets developed by the European Committee for Standardization (EN). <sup>c</sup> Na<sub>2</sub>C<sub>6</sub>H<sub>8</sub>O<sub>8</sub> = sodium hydrogen citrate sesquihydrate.

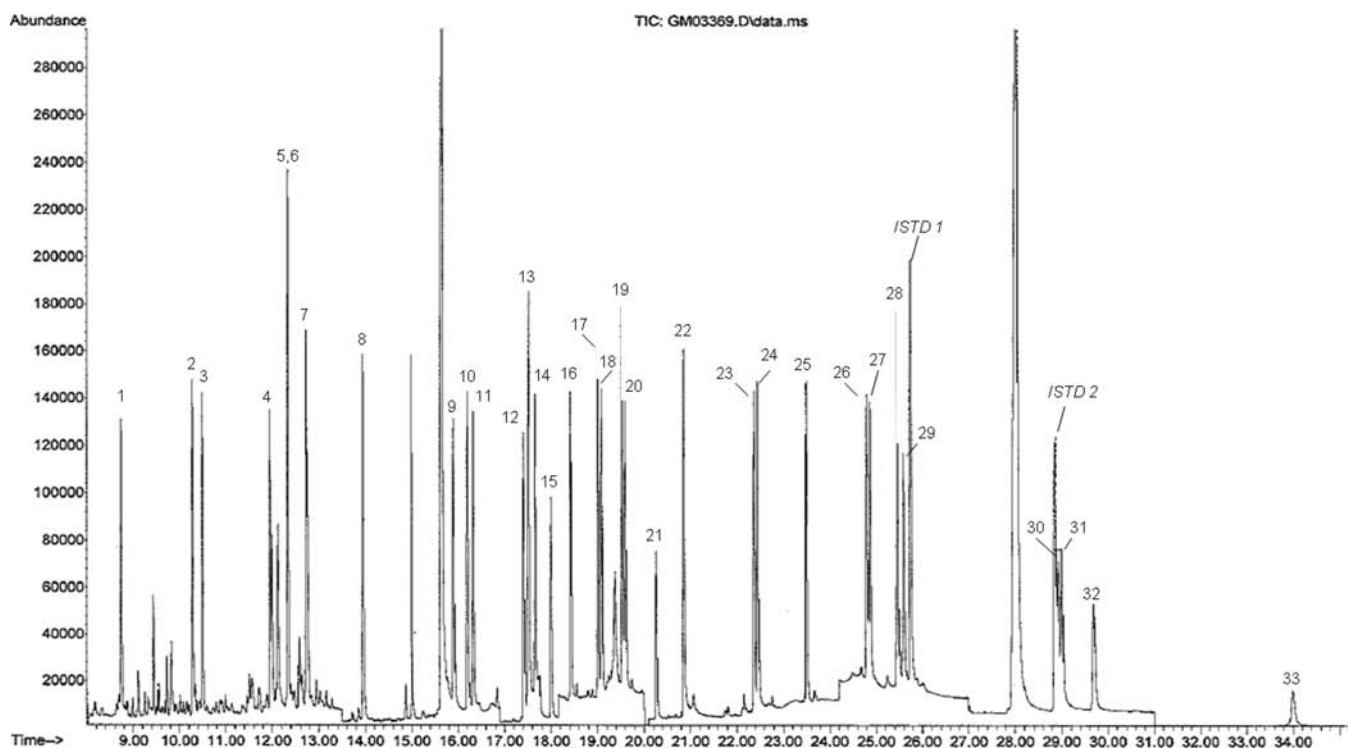


Figure 1. Representative selective ion monitoring total ion current (TIC) for PAHs in smoked salmon with a 500 ng/g wet weight overspike. PAHs corresponding to chromatogram numbers can be found in Table 1.

**GC–MS Analysis.** All standards and samples were analyzed using an Agilent 5975B GC–MS (Santa Clara, CA) with electron impact ionization (70 eV) utilizing selective ion monitoring (SIM) in positive ion mode and a DB-5MS column (30 m length, 0.25  $\mu$ m film thickness, 0.25 mm i.d., Agilent J&W). The instrument injection port was operated in the pulsed splitless mode, fitted with a 2 mm glass liner with deactivated glass wool, and delivered a 1  $\mu$ L injection to an inlet maintained at 300 °C. Chromatography of PAHs was achieved using the following program at a column flow rate of 1 mL/min using helium as the carrier gas: the initial oven temperature was 70 °C, 1 min hold, ramp to 300 at 10 °C/min, 4 min hold, ramp to 310 at 10 °C/min, 7 min hold for a total run time of 36 min. Mass spectrometer transfer line, source and quadrupole temperatures were 280 °C, 230 °C and 150 °C respectively. PAH SIM ions are presented in Table 1 along with retention times and coefficients of determination.

**Sample Preparation and Fortification.** Sample preparation followed a modified QuEChERS methodology.<sup>14</sup> For PAH spike and recovery experiments, 1 g of salmon homogenate (wet weight) was weighed into a 15 mL conical centrifuge tube using a Brinkmann Instruments Inc.

Sartorius 1202 MP analytical balance (Westbury, NY) and allowed to come to room temperature. Spiked samples were fortified with 50  $\mu$ L of a 10  $\mu$ g/mL composite of 33 PAHs (500 ng/g) directly onto the fish matrix and allowed to acclimate for 2 min. Matrix blanks were prepared similarly but were not fortified. Samples were then extracted by one of four methods.

#### Sample Extraction Procedures (See Table 2 for Summary).

Four extraction methods (E1, E2, E3 and E4) were used in the study. Schemes E1 and E2 represent methods commonly referred to as the “traditional QuEChERS methods” and employ AOAC and EN salts respectively. Full details for extraction schemes E1 and E2 are described elsewhere.<sup>20,22–24</sup> Briefly, E1 samples received 4 mL of H<sub>2</sub>O followed by vigorous shaking/vortexing for 1 min. To this slurry was added 5 mL of 1% acetic acid in acetonitrile, and the resulting mixture was mixed for 1 min. Next, 2.5 g of AOAC extraction salts were added and the mixture was shaken/vortexed for 1 min and centrifuged at 3800g for 5 min with an Eppendorf 5810R centrifuge (Westbury, NY). E2 samples received 1 mL of H<sub>2</sub>O and were shaken/vortexed for 1 min. Then 2 mL of acetonitrile was added and the resulting mixture was mixed thoroughly

**Table 3. Traditional QuEChERS Method Performance in Recovering 33 PAHs (Mean  $\pm$  RSD;  $n = 4$ ) from Increasing Fat Content Smoked Salmon Fortified at 500 ng/g Wet Weight**

PAH	recovery (%)					
	E1 <sup>a</sup>			E2 <sup>a</sup>		
	3% <sup>b</sup>	8%	11%	3%	8%	11%
naphthalene	39.6 $\pm$ 47.7	30.0 $\pm$ 5.5	45.5 $\pm$ 8.4	34.3 $\pm$ 45.6	38.7 $\pm$ 5.8	43.3 $\pm$ 12.7
2-methylnaphthalene	48.8 $\pm$ 32.2	42.5 $\pm$ 2.2	53.2 $\pm$ 5.3	42.2 $\pm$ 35.1	47.5 $\pm$ 5.0	49.6 $\pm$ 10.4
1-methylnaphthalene	47.0 $\pm$ 29.5	44.1 $\pm$ 0.9	54.2 $\pm$ 5.7	43.1 $\pm$ 31.9	47.5 $\pm$ 3.0	50.3 $\pm$ 9.5
1,6-dimethylnaphthalene	55.0 $\pm$ 21.9	51.2 $\pm$ 0.5	58.5 $\pm$ 3.9	48.8 $\pm$ 28.2	52.0 $\pm$ 3.6	54.2 $\pm$ 7.8
1,2-dimethylnaphthalene	62.3 $\pm$ 26.5	61.4 $\pm$ 3.8	60.7 $\pm$ 3.1	49.7 $\pm$ 26.8	56.6 $\pm$ 1.8	55.4 $\pm$ 7.8
acenaphthylene	56.5 $\pm$ 21.9	57.1 $\pm$ 2.1	58.2 $\pm$ 4.5	50.2 $\pm$ 25.0	58.0 $\pm$ 2.3	55.6 $\pm$ 6.9
acenaphthene	56.1 $\pm$ 21.2	55.0 $\pm$ 1.5	56.9 $\pm$ 3.7	47.8 $\pm$ 25.5	55.1 $\pm$ 1.7	54.0 $\pm$ 7.5
fluorene	60.6 $\pm$ 18.5	62.7 $\pm$ 1.5	65.0 $\pm$ 3.6	54.5 $\pm$ 22.0	64.6 $\pm$ 1.7	61.8 $\pm$ 5.6
dibenzothiophene	62.7 $\pm$ 12.7	68.1 $\pm$ 2.1	62.8 $\pm$ 2.7	58.9 $\pm$ 15.0	71.4 $\pm$ 1.6	62.4 $\pm$ 4.0
phenanthrene	65.4 $\pm$ 13.4	74.2 $\pm$ 2.0	66.8 $\pm$ 2.3	63.0 $\pm$ 12.2	76.0 $\pm$ 2.0	66.0 $\pm$ 3.5
anthracene	71.0 $\pm$ 10.2	82.0 $\pm$ 1.6	73.6 $\pm$ 2.5	67.1 $\pm$ 13.3	82.4 $\pm$ 2.0	70.3 $\pm$ 3.6
2-methylphenanthrene	73.0 $\pm$ 8.1	83.2 $\pm$ 1.8	70.8 $\pm$ 3.0	71.5 $\pm$ 8.6	85.1 $\pm$ 1.6	71.9 $\pm$ 2.7
2-methylanthracene	71.1 $\pm$ 10.1	66.3 $\pm$ 44.5	33.9 $\pm$ 5.3	73.4 $\pm$ 11.8	93.5 $\pm$ 2.3	74.4 $\pm$ 3.8
1-methylphenanthrene	72.8 $\pm$ 8.8	84.6 $\pm$ 1.0	69.0 $\pm$ 3.7	66.9 $\pm$ 8.4	79.3 $\pm$ 1.1	68.3 $\pm$ 2.5
9-methylanthracene	76.3 $\pm$ 6.2	83.7 $\pm$ 2.2	73.2 $\pm$ 1.9	70.8 $\pm$ 8.5	83.6 $\pm$ 1.3	70.0 $\pm$ 2.6
3,6-dimethylphenanthrene	74.0 $\pm$ 5.2	77.0 $\pm$ 1.4	68.4 $\pm$ 2.5	68.6 $\pm$ 7.9	77.9 $\pm$ 1.3	66.4 $\pm$ 2.5
fluoranthene	75.0 $\pm$ 4.2	84.4 $\pm$ 2.1	69.6 $\pm$ 2.4	72.1 $\pm$ 5.3	85.3 $\pm$ 0.4	69.1 $\pm$ 1.3
2,3-dimethylanthracene	74.1 $\pm$ 3.8	79.8 $\pm$ 1.8	67.8 $\pm$ 2.6	70.7 $\pm$ 6.2	80.3 $\pm$ 1.0	66.2 $\pm$ 2.1
9,10-dimethylanthracene	76.7 $\pm$ 3.3	79.3 $\pm$ 2.1	70.6 $\pm$ 2.4	72.4 $\pm$ 5.1	79.0 $\pm$ 0.8	66.7 $\pm$ 1.8
pyrene	73.2 $\pm$ 4.5	83.4 $\pm$ 2.7	67.1 $\pm$ 1.9	71.1 $\pm$ 4.5	82.1 $\pm$ 0.5	65.8 $\pm$ 1.4
retene	73.1 $\pm$ 2.9	74.5 $\pm$ 2.0	66.1 $\pm$ 3.6	69.4 $\pm$ 5.2	76.0 $\pm$ 1.1	64.4 $\pm$ 1.7
1-methylpyrene	69.2 $\pm$ 2.5	78.4 $\pm$ 1.9	65.2 $\pm$ 7.6	66.8 $\pm$ 3.7	77.0 $\pm$ 0.5	63.9 $\pm$ 1.5
benz[ <i>a</i> ]anthracene	87.6 $\pm$ 3.8	96.0 $\pm$ 2.0	80.0 $\pm$ 2.7	88.9 $\pm$ 2.8	95.9 $\pm$ 0.4	80.4 $\pm$ 0.7
chrysene	75.9 $\pm$ 3.3	83.6 $\pm$ 2.2	69.5 $\pm$ 2.7	77.4 $\pm$ 2.8	83.0 $\pm$ 0.6	69.7 $\pm$ 0.8
6-methylchrysene	73.2 $\pm$ 2.0	72.5 $\pm$ 2.0	62.8 $\pm$ 2.2	71.7 $\pm$ 3.8	71.9 $\pm$ 0.6	62.9 $\pm$ 0.9
benzo[ <i>b</i> ]fluoranthene	68.6 $\pm$ 3.8	70.8 $\pm$ 2.3	61.4 $\pm$ 2.9	73.4 $\pm$ 2.4	70.9 $\pm$ 1.1	63.6 $\pm$ 0.8
benzo[ <i>k</i> ]fluoranthene	66.2 $\pm$ 4.5	68.4 $\pm$ 2.0	59.3 $\pm$ 3.8	72.6 $\pm$ 2.6	69.1 $\pm$ 0.9	62.2 $\pm$ 1.1
benzo[ <i>e</i> ]pyrene	66.4 $\pm$ 3.1	65.3 $\pm$ 2.1	57.1 $\pm$ 2.8	65.1 $\pm$ 2.3	63.2 $\pm$ 0.8	56.6 $\pm$ 1.2
benzo[ <i>a</i> ]pyrene	64.0 $\pm$ 4.4	63.8 $\pm$ 2.0	55.6 $\pm$ 3.0	67.0 $\pm$ 2.5	60.8 $\pm$ 1.0	56.4 $\pm$ 1.5
indeno[1,2,3- <i>cd</i> ]pyrene	53.2 $\pm$ 4.0	54.6 $\pm$ 1.3	47.6 $\pm$ 2.5	62.0 $\pm$ 1.6	53.8 $\pm$ 1.1	51.3 $\pm$ 1.6
dibenz[ <i>a,h</i> ]anthracene	63.5 $\pm$ 2.9	66.6 $\pm$ 0.5	55.6 $\pm$ 2.6	68.9 $\pm$ 1.2	62.7 $\pm$ 0.6	59.0 $\pm$ 1.4
benzo[ <i>g,h,i</i> ]perylene	51.7 $\pm$ 2.2	47.7 $\pm$ 1.6	44.0 $\pm$ 2.7	56.7 $\pm$ 1.5	47.2 $\pm$ 0.4	46.2 $\pm$ 2.0
dibenzo[ <i>a,l</i> ]pyrene	62.6 $\pm$ 4.5	34.2 $\pm$ 4.2	45.0 $\pm$ 8.7	71.5 $\pm$ 1.3	24.5 $\pm$ 8.6	46.7 $\pm$ 2.0
av recovery across all PAHs (%)	66	67	61	64	68	61

<sup>a</sup> See Table 2 for method specifics. <sup>b</sup> Fat content.

for 15 min. Samples were subsequently treated with 1.3 g of EN extraction salts, shaken/vortexed for 15 min and centrifuged at 3800g for 5 min.

Two new extraction schemes, E3 and E4, were developed as modifications to E1 and E2. Scheme E3 and E4 samples received 1 mL of H<sub>2</sub>O followed by vigorous shaking/vortexing for 1 min. The resulting slurries were treated with 2 mL of a solution of high purity acetone, ethyl acetate and isooctane (2:2:1; v/v/v) and thoroughly mixed for 5 min. Then, samples were treated with 1.3 g of either AOAC or EN extraction salts, shaken/vortexed for 5 min and centrifuged at 3800g for 5 min. Samples treated with AOAC and EN salts are denoted as schemes E3 and E4 respectively.

**Sample Cleanup and Internal Standard Addition Procedure.** All extracted samples were subject to dispersive solid-phase extraction as it has been previously demonstrated to improve PAH recoveries in shrimp and sample drying.<sup>14,21</sup> Extracts (1 mL) were aliquoted

into commercially available 2 mL Sampli-Q AOAC fatty sample dispersive SPE tubes, shaken/vortexed for 5 min and centrifuged at 13600g for 5 min with an Eppendorf 5415C microcentrifuge (Westbury, NY). Aliquots of the resulting supernatant (200  $\mu$ L) were transferred to autosampler vials fitted with small volume inserts, spiked with perylene-D12 and indeno[1,2,3-*cd*]pyrene-D12 internal standards, vortex mixed and stored in the dark at  $-20^{\circ}$ C until analysis. All spike-recovery experiments and matrix blank determinations were conducted in replicates of four and three respectively.

**PAH and Substituted PAH Quantification.** Following extraction, cleanup and internal standard addition, all samples were quantified for PAHs and substituted PAHs using GC-MS. Purchased native standards were used to accurately identify and quantify PAHs and their substituted derivatives. Analyte concentrations were determined from calibration curves of relative response factors of analytes to internal

**Table 4. Modified QuEChERS Method Performance in Recovering 33 PAHs (Mean  $\pm$  RSD;  $n = 4$ ) from Increasing Fat Content Smoked Salmon Fortified at 500 ng/g Wet Weight**

PAH	recovery (%)					
	E3 <sup>a</sup>			E4 <sup>a</sup>		
	3% <sup>b</sup>	8%	11%	3%	8%	11%
naphthalene	72.4 $\pm$ 2.5	98.3 $\pm$ 3.4	76.4 $\pm$ 1.6	73.4 $\pm$ 7.0	96.0 $\pm$ 3.1	70.0 $\pm$ 3.4
2-methylnaphthalene	77.5 $\pm$ 3.1	102.7 $\pm$ 2.5	82.3 $\pm$ 1.8	79.1 $\pm$ 7.4	96.5 $\pm$ 2.2	76.0 $\pm$ 2.7
1-methylnaphthalene	73.8 $\pm$ 3.5	98.7 $\pm$ 2.7	80.5 $\pm$ 2.5	75.6 $\pm$ 7.3	94.6 $\pm$ 3.7	73.6 $\pm$ 2.1
1,6-dimethylnaphthalene	80.9 $\pm$ 3.5	107.4 $\pm$ 2.6	85.6 $\pm$ 1.6	83.2 $\pm$ 7.0	102.1 $\pm$ 3.7	77.9 $\pm$ 2.0
1,2-dimethylnaphthalene	81.1 $\pm$ 2.4	108.0 $\pm$ 3.1	81.2 $\pm$ 2.2	82.9 $\pm$ 6.1	102.3 $\pm$ 3.4	74.6 $\pm$ 2.2
acenaphthylene	77.7 $\pm$ 3.6	103.0 $\pm$ 2.5	80.3 $\pm$ 1.3	80.0 $\pm$ 6.9	98.0 $\pm$ 3.5	73.1 $\pm$ 1.7
acenaphthene	76.8 $\pm$ 3.5	103.4 $\pm$ 2.6	79.2 $\pm$ 1.4	78.6 $\pm$ 7.0	97.4 $\pm$ 3.2	71.3 $\pm$ 1.8
fluorene	81.9 $\pm$ 3.7	107.9 $\pm$ 2.8	86.9 $\pm$ 1.5	84.1 $\pm$ 7.0	102.2 $\pm$ 3.7	79.8 $\pm$ 1.9
dibenzothiophene	79.2 $\pm$ 3.8	105.2 $\pm$ 2.4	82.6 $\pm$ 1.5	82.3 $\pm$ 6.9	100.1 $\pm$ 4.1	76.9 $\pm$ 1.8
phenanthrene	80.5 $\pm$ 3.8	107.8 $\pm$ 2.8	85.1 $\pm$ 1.6	82.3 $\pm$ 7.2	101.9 $\pm$ 3.8	78.4 $\pm$ 2.0
anthracene	89.9 $\pm$ 4.2	120.4 $\pm$ 2.8	95.4 $\pm$ 1.6	92.7 $\pm$ 7.2	113.5 $\pm$ 4.0	87.8 $\pm$ 2.0
2-methylphenanthrene	88.2 $\pm$ 4.0	122.0 $\pm$ 2.8	88.8 $\pm$ 12.0	90.9 $\pm$ 6.8	113.7 $\pm$ 3.9	85.8 $\pm$ 2.1
2-methylanthracene	76.6 $\pm$ 6.8	128.7 $\pm$ 12.5	84.5 $\pm$ 13.4	93.7 $\pm$ 4.0	75.1 $\pm$ 5.4	79.9 $\pm$ 2.4
1-methylphenanthrene	85.0 $\pm$ 3.9	115.1 $\pm$ 1.8	92.5 $\pm$ 1.5	88.6 $\pm$ 8.2	111.2 $\pm$ 3.6	86.2 $\pm$ 2.4
9-methylanthracene	90.7 $\pm$ 3.9	117.4 $\pm$ 2.4	95.6 $\pm$ 1.4	92.8 $\pm$ 7.4	110.7 $\pm$ 3.7	87.9 $\pm$ 1.5
3,6-dimethylphenanthrene	85.3 $\pm$ 3.2	108.9 $\pm$ 2.6	89.2 $\pm$ 1.5	88.1 $\pm$ 7.8	104.3 $\pm$ 3.7	84.5 $\pm$ 1.7
fluoranthene	84.6 $\pm$ 3.6	111.9 $\pm$ 2.7	88.5 $\pm$ 2.0	86.4 $\pm$ 6.9	105.5 $\pm$ 3.6	81.1 $\pm$ 1.9
2,3-dimethylanthracene	91.4 $\pm$ 3.7	115.4 $\pm$ 2.8	94.9 $\pm$ 1.8	93.7 $\pm$ 6.9	108.3 $\pm$ 3.9	88.9 $\pm$ 1.7
9,10-dimethylanthracene	89.9 $\pm$ 3.3	113.6 $\pm$ 2.3	94.6 $\pm$ 1.6	91.8 $\pm$ 7.0	107.4 $\pm$ 3.8	87.8 $\pm$ 1.9
pyrene	84.2 $\pm$ 3.8	117.0 $\pm$ 2.6	87.8 $\pm$ 3.3	86.2 $\pm$ 6.7	106.0 $\pm$ 3.1	79.9 $\pm$ 2.4
retene	90.1 $\pm$ 3.3	112.9 $\pm$ 2.5	97.0 $\pm$ 1.7	92.2 $\pm$ 6.4	106.4 $\pm$ 3.7	90.2 $\pm$ 1.8
1-methylpyrene	83.5 $\pm$ 3.3	106.8 $\pm$ 2.4	92.3 $\pm$ 1.7	84.2 $\pm$ 7.3	100.6 $\pm$ 3.5	83.2 $\pm$ 1.7
benz[ <i>a</i> ]anthracene	102.2 $\pm$ 3.9	130.4 $\pm$ 2.6	106.0 $\pm$ 1.5	105.1 $\pm$ 6.9	122.6 $\pm$ 4.0	98.1 $\pm$ 1.9
chrysene	89.8 $\pm$ 3.8	114.1 $\pm$ 2.8	92.9 $\pm$ 1.6	92.1 $\pm$ 6.9	107.1 $\pm$ 3.9	86.0 $\pm$ 1.8
6-methylchrysene	82.7 $\pm$ 2.8	98.9 $\pm$ 2.4	84.6 $\pm$ 1.5	84.5 $\pm$ 7.5	93.6 $\pm$ 3.7	79.3 $\pm$ 2.1
benzo[ <i>b</i> ]fluoranthene	87.2 $\pm$ 3.9	97.1 $\pm$ 1.9	85.9 $\pm$ 1.6	89.5 $\pm$ 6.3	92.4 $\pm$ 3.8	80.3 $\pm$ 2.2
benzo[ <i>k</i> ]fluoranthene	87.9 $\pm$ 4.2	96.7 $\pm$ 0.7	85.7 $\pm$ 1.6	89.6 $\pm$ 5.8	92.4 $\pm$ 4.1	79.8 $\pm$ 2.6
benzo[ <i>e</i> ]pyrene	78.3 $\pm$ 3.0	83.4 $\pm$ 1.6	75.4 $\pm$ 1.0	79.6 $\pm$ 6.3	80.7 $\pm$ 2.8	70.3 $\pm$ 1.4
benzo[ <i>a</i> ]pyrene	85.3 $\pm$ 3.3	89.2 $\pm$ 1.7	82.0 $\pm$ 1.0	87.4 $\pm$ 6.7	85.4 $\pm$ 3.2	76.2 $\pm$ 1.2
indeno[1,2,3- <i>cd</i> ]pyrene	80.1 $\pm$ 2.3	82.5 $\pm$ 2.6	79.6 $\pm$ 2.5	83.5 $\pm$ 6.1	77.9 $\pm$ 4.4	72.9 $\pm$ 1.1
dibenzo[ <i>a,h</i> ]anthracene	83.2 $\pm$ 2.5	84.0 $\pm$ 2.3	81.4 $\pm$ 1.4	86.6 $\pm$ 6.3	79.6 $\pm$ 4.3	74.9 $\pm$ 1.3
benzo[ <i>g,h,i</i> ]perylene	74.1 $\pm$ 1.7	69.5 $\pm$ 2.9	68.1 $\pm$ 1.9	76.5 $\pm$ 5.1	67.2 $\pm$ 3.6	61.9 $\pm$ 0.4
dibenzo[ <i>a,l</i> ]pyrene	67.6 $\pm$ 2.6	40.5 $\pm$ 10.1	48.5 $\pm$ 3.5	77.5 $\pm$ 4.9	52.4 $\pm$ 4.4	36.0 $\pm$ 3.0
av recovery across all PAHs (%)	83	104	85	86	97	78

<sup>a</sup> See Table 2 for method specifics. <sup>b</sup> Fat content.

standards. Calibration curves were generated from seven calibration standards prepared in isoctane with a concentration range of 1–1000 pg/ $\mu$ L. Extractions of nonspiked salmon matrix were performed in replicate ( $n = 3$ ) for PAH background determination. Method recoveries (%) were subsequently background subtracted.

**Quality Assurance/Control.** Each analytical batch contained a minimum of 15% quality control samples, including solvent blanks, check standards and overspikes. Instrument stability was assessed by analyzing continuing calibration verification standards every 6–8 samples. The accuracy and precision of continuing calibration verification standards were typically within  $\pm 15\%$  of expected values. Additionally, interday/batch accuracy and precision over four analytical batches run on four different days were typically within  $\pm 10\%$  of expected values. Finally, the presence of artifact PAHs arising from laboratory associated procedures was assessed through the analysis of laboratory reagent blanks. Parent

and substituted PAH residues were not detected in any laboratory reagent blank samples.

## RESULTS AND DISCUSSION

Figure 1 is an example chromatogram generated from salmon spike-recovery experiments demonstrating sufficient separation/detection of 33 PAHs and substituted PAHs by GC–MS in 36 min at a sample overspike concentration of 500 ng/g (wet weight). Table 1 summarizes native PAH and deuterated internal standard GC–MS instrumental parameters and detection limits. Benz[*a*]anthracene was the only compound that had a coefficient of determination ( $r^2$ ) less than 0.99 ( $r^2 = 0.983$ ); all others had coefficients  $\geq 0.99$  within the calibration range of 1–1000 pg/ $\mu$ L, demonstrating excellent method linearity. Parent and substituted

**Table 5. Modified QuEChERS extraction performance (% recovery  $\pm$  SD) compared to literature reported Soxhlet and accelerated solvent extraction (ASE) in recovering 15 PAHs from fish tissues**

PAH	recovery (%)				
	Soxhlet with hexane <sup>a</sup>	ASE with hexane <sup>a</sup>	ASE with CH <sub>2</sub> Cl <sub>2</sub> :ACN (9:1) <sup>a</sup>	modified QuEChERS E3 <sup>b</sup>	modified QuEChERS E4 <sup>c</sup>
naphthalene	51 $\pm$ 6	58 $\pm$ 5	62 $\pm$ 6	82 $\pm$ 12	80 $\pm$ 13
acenaphthylene	84 $\pm$ 7	88 $\pm$ 9	76 $\pm$ 5	87 $\pm$ 12	84 $\pm$ 12
acenaphthene	68 $\pm$ 4	67 $\pm$ 6	69 $\pm$ 4	86 $\pm$ 13	82 $\pm$ 12
fluorene	77 $\pm$ 6	81 $\pm$ 7	77 $\pm$ 10	92 $\pm$ 12	89 $\pm$ 11
phenanthrene	91 $\pm$ 12	71 $\pm$ 11	93 $\pm$ 8	91 $\pm$ 13	88 $\pm$ 11
anthracene	66 $\pm$ 5	61 $\pm$ 5	81 $\pm$ 7	102 $\pm$ 14	98 $\pm$ 12
fluoranthene	71 $\pm$ 9	73 $\pm$ 9	101 $\pm$ 7	95 $\pm$ 13	91 $\pm$ 12
pyrene	69 $\pm$ 7	68 $\pm$ 6	92 $\pm$ 5	96 $\pm$ 16	91 $\pm$ 12
benz[ <i>a</i> ]anthracene	70 $\pm$ 5	68 $\pm$ 4	96 $\pm$ 11	113 $\pm$ 13	109 $\pm$ 12
chrysene	44 $\pm$ 3	53 $\pm$ 4	93 $\pm$ 9	99 $\pm$ 12	95 $\pm$ 10
benzo[ <i>k</i> ]fluoranthene	45 $\pm$ 10	54 $\pm$ 11	89 $\pm$ 5	90 $\pm$ 5	87 $\pm$ 7
benzo[ <i>a</i> ]pyrene	74 $\pm$ 14	57 $\pm$ 6	79 $\pm$ 8	85 $\pm$ 4	83 $\pm$ 6
indeno[1,2,3- <i>cd</i> ]pyrene	56 $\pm$ 6	56 $\pm$ 12	81 $\pm$ 3	81 $\pm$ 2	78 $\pm$ 6
dibenzo[ <i>a,h</i> ]anthracene	55 $\pm$ 8	48 $\pm$ 8	84 $\pm$ 8	83 $\pm$ 2	80 $\pm$ 6
benzo[ <i>g,h,i</i> ]perylene	61 $\pm$ 11	67 $\pm$ 10	85 $\pm$ 7	71 $\pm$ 3	69 $\pm$ 7
av recovery across all PAHs (%)	65	65	84	90	87

<sup>a</sup> Values originally reported by Wang et al. (1999);  $n = 4$  replicates for each extraction method. <sup>b</sup> Values represent mean recovery across all three fat level fish tested by extraction method E3;  $n = 12$ . <sup>c</sup> Values represent mean recovery across all three fat level fish tested by extraction method E4;  $n = 12$ .

PAH instrumental detection limits were assigned to PAH molecular ions when their lowest abundance confirmation ion signal-to-noise (S/N)  $\geq 3$  as determined by the signal-to-noise script of the Agilent MSD ChemStation data analysis software, version E (Santa Clara, CA). Samples used in the determination of instrumental detection limits were standard solutions analyzed from several batches over several days. The instrumental detection limits for quantified analytes ranged from 1 to 5 pg/ $\mu$ L. Analytes were considered quantitative when they calibrated with  $r^2 \geq 0.98$ , their lowest abundance confirmation ion had S/N  $> 3$  and they had reproducible and accurate quantitation ( $\pm 20\%$  of their true value) as assessed from continuing calibration verification standards. All parent and substituted PAHs met these criteria. Sample residues that met all criteria but had S/N  $< 3$  were designated below detection limit (BDL), while those that did not meet one or more of the above criteria were designated nondetectable (ND).

Table 3 summarizes PAH spike recoveries obtained for traditional acetonitrile based QuEChERS extraction methods E1 and E2 from smoked salmon with 3, 8, and 11% fat content. Recoveries from smoked salmon using extraction scheme E1 (1% acetic acid in acetonitrile and AOAC salts) yielded low recoveries, on average less than 67%, with individual PAH recoveries typically ranging from 35 to 87%. Extraction scheme E2 (acetonitrile and EN salts) performed equally poorly, with average PAH recoveries being less than 68% and individual PAH recoveries ranging from 24 to 88%. Both extraction schemes E1 and E2 were especially poor at recovering 2-, 3-, 5- and 6-ring PAHs, where average recoveries across this subgroup of PAHs were 57% and 56% respectively.

Variants QuEChERS solvent systems have been described for the analysis of pesticide residues in fruits.<sup>14,28</sup> Additionally, the individual and combined performance of various ratios of ethyl acetate, acetone, hexane, methylene chloride, acetonitrile, cyclohexane and isooctane have been reported for multiple residue

pesticide methods and EPA methods for extraction of nonvolatile and semivolatile organic compounds from solid and semisolid samples.<sup>10,12,13,28</sup> Of interest were solvent systems with improved selectivity for nonpolar PAH residues and that were lower in cost than acetonitrile. It was found that a three-component variant solvent system of acetone, ethyl acetate and isooctane (2:2:1; v/v/v) met these criteria.

Table 4 summarizes PAH recoveries obtained from smoked salmon using modified QuEChERS extraction schemes E3 and E4. Extraction scheme E3 (acetone, ethyl acetate, isooctane and AOAC salts) led to good recoveries, on average 90% over all fish tested. Notable performance gains were made for 2-, 3- and 5-ring PAHs where recoveries were improved 50–200%, while recoveries of 4- and 6-ring PAHs were slightly improved by ~30–45% as compared to acetonitrile. Extraction scheme E4 (acetone, ethyl acetate, isooctane and EN salts) performed equally well, with an average PAH recovery of 87% across all fish and individual PAHs displaying the same range of improvement as extraction scheme E3. Additionally, both extraction schemes E3 and E4 displayed good extraction precision with relative standard deviations typically less than 10% for all fish tested.

It is well understood that the planar hydrophobic chemical structure of PAHs leads to their association with fatty components of biological matrices (i.e., waxes, lipids, steroids and pigments). Extraction conditions that disrupt these associations should give rise to enhanced extraction performance. A solvent's ability to disrupt interactions may be assessed by comparison of solvent and PAH octanol–water partition coefficients (log  $K_{OW}$ ), where solvents with coefficients similar to PAHs should display enhanced selectivity. The log  $K_{OW}$  for PAHs used in this study ranged from 3.3 for naphthalene to 7.7 for dibenzo[*a,l*]pyrene. Acetonitrile has a reported log  $K_{OW} = -0.34$ , while values for acetone, ethyl acetate and isooctane are  $-0.24$ ,  $0.73$ , and  $4.1$  respectively. It has also been demonstrated that extraction of various food stuffs with acetonitrile resulted in limited

extraction of hydrophobic matrix components.<sup>14,29</sup> This information coupled to our results suggests that the three component extraction solvent used in extraction schemes E3 and E4 possesses physicochemical characteristics that allow it to interact more intimately with fatty fish matrices. At the molecular level, extraction with acetone, ethyl acetate and isooctane may lead to improved recoveries by allowing water miscible acetone and ethyl acetate to recover PAHs trapped in water-sealed matrix pores and making them available for transfer to isooctane.

It is also known that increased extraction temperatures can disrupt analyte–matrix interactions by decreasing the activation energy required for analyte desorption processes and decreasing solvent viscosity, facilitating better solvent–matrix penetration.<sup>12</sup> It was found that addition of magnesium sulfate containing extraction salts generated sample extraction temperatures of 45–50 °C that persisted for the duration of the extraction/partition procedure (data not shown). Extraction temperatures in the range observed have been reported by others and should increase solvent capacity for PAHs.<sup>12,14</sup> The findings presented indicate that the improved extraction performance of schemes E3 and E4 likely resulted from the combined influence of enhanced solvent selectivity and elevated sample extraction temperatures.

In order to evaluate the effectiveness and utility of the modified QuEChERS methods developed in this study, a comparison to other published extraction techniques is presented (Table 5).<sup>13</sup> Compared to Soxhlet extraction with hexane, it was found that modified QuEChERS methods substantially improved average recovery of all 15 PAHs by roughly 38% and led to individual gains of 50–125% for naphthalene, anthracene, benz[*a*]anthracene, benzo[*k*]fluoranthene and dibenz[*a,h*]anthracene. Similar overall improvements were demonstrated when compared to accelerated solvent extraction (ASE) with hexane. Interestingly, the performance of modified QuEChERS methods was comparable to a validated dichloromethane and acetonitrile (1:1; v/v) based ASE extraction method in average recovery across all PAHs. However, modified QuEChERS methods showed improved recoveries in lower molecular weight PAHs, while ASE performed better at recovering benzo[*g,h,i*]perylene. Estimated method detection limits (MDL) for modified QuEChERS methods are presented in Table 6 in relation to FDA PAH levels of concern in shrimp, crab, oysters and finfish.<sup>25</sup> MDLs were defined as the product of the analyte instrument detection and the method dilution factor, which was a factor of 2 in this case. MDLs were all well below levels of concern in these food stuffs, demonstrating the potential utility of the developed methods. If needed, additional method sensitivity could be achieved through the introduction of a solvent reduction procedure prior to instrumental analysis.

A comparison of PAH levels measured in commercially available smoked salmon using both modified QuEChERS extraction schemes E3 and E4 is presented in Table 7. Results for the two methods were quite similar across all fish in terms of PAH profile, quantitation, extraction precision and sum of quantified PAHs ( $\Sigma$ PAHs). Naphthalene, 2-methylnaphthalene, 1-methylnaphthalene, phenanthrene, fluoranthene, pyrene and benzo[*g,h,i*]perylene were consistently detected. Anthracene was observed only in 3% fat salmon, but was not quantified because it did not meet signal-to-noise limits. Levels of individual PAHs typically fell in a range of 5–60 ng/g wet weight with fluoranthene and benzo[*g,h,i*]perylene displaying the lowest levels and naphthalene, 2-methylnaphthalene and pyrene consistently accounting for >70% of total PAH mass recovered. Extraction schemes E3 and E4 also performed equally well with regard to extraction precision

**Table 6. Comparison of FDA PAH Levels of Concern in Relation to Estimated Modified QuEChERS Method Detection Limits (MDL)**

PAH	FDA levels of concern ( $\mu\text{g/g}$ )			modified QuEChERS MDL ( $\mu\text{g/g}$ ) <sup>b</sup>
	shrimp and crab <sup>a</sup>	oyster <sup>a</sup>	finfish <sup>a</sup>	
naphthalene	123	133	32.7	0.002
2-methylnaphthalene	NA <sup>c</sup>	NA	NA	0.002
1-methylnaphthalene	NA	NA	NA	0.002
1,6-dimethylnaphthalene	NA	NA	NA	0.002
acenaphthylene	NA	NA	NA	0.002
1,2-dimethylnaphthalene	NA	NA	NA	0.002
acenaphthene	NA	NA	NA	0.002
fluorene	246	267	65.3	0.002
dibenzothiophene	NA	NA	NA	0.002
phenanthrene	1846 <sup>d</sup>	2000 <sup>d</sup>	490 <sup>d</sup>	0.010
anthracene	NA	NA	NA	0.010
2-methylphenanthrene	NA	NA	NA	0.010
2-methylanthracene	NA	NA	NA	0.010
1-methylphenanthrene	NA	NA	NA	0.010
9-methylanthracene	NA	NA	NA	0.010
3,6-dimethylphenanthrene	NA	NA	NA	0.002
fluoranthene	246	267	65.3	0.002
2,3-dimethylanthracene	NA	NA	NA	0.010
pyrene	185	200	49.0	0.002
9,10-dimethylanthracene	NA	NA	NA	0.002
retene	NA	NA	NA	0.002
1-methylpyrene	NA	NA	NA	0.002
benz[ <i>a</i> ]anthracene	1.32	1.43	0.35	0.010
chrysene	132	143	35.0	0.002
6-methylchrysene	NA	NA	NA	0.002
benzo[ <i>b</i> ]fluoranthene	1.32	1.43	0.35	0.002
benzo[ <i>k</i> ]fluoranthene	13.2	14.3	3.5	0.002
benzo[ <i>e</i> ]pyrene	NA	NA	NA	0.002
benzo[ <i>a</i> ]pyrene	0.132	0.143	0.035	0.002
indeno[1,2,3- <i>cd</i> ]pyrene	1.32	1.43	0.35	0.002
dibenz[ <i>a,h</i> ]anthracene	0.132	0.143	0.035	0.010
benzo[ <i>g,h,i</i> ]perylene	NA	NA	NA	0.002
dibenzo[ <i>a,l</i> ]pyrene	NA	NA	NA	0.002

<sup>a</sup> Values obtained from U.S. FDA (2010). <sup>b</sup> MDL = IDL multiplied by a dilution factor of 2. <sup>c</sup> Levels of concern not known at time of publication. <sup>d</sup> Represents summed levels of concern for phenanthrene and anthracene.

(RSD < 20%) and produced nearly identical  $\Sigma$ PAH values within and across the fat levels used. Results from this study are comparable to those from other studies in terms of PAH profile, range and summed residue loads and, together with spike-recovery experiments, demonstrate that choice of extraction solvent is crucial to extraction performance.<sup>6,7,13</sup>

The goal of the current study was to develop and validate a multiresidue method for the analysis of PAHs and their substituted derivatives in high-fat smoked salmon. The data presented strongly indicate that a QuEChERS based analytical

**Table 7. Comparison of PAH Levels (Mean ng/g Wet Weight  $\pm$  SD,  $n = 3$ ) Measured in Commercially Available Smoked Salmon by Two New Modified QuEChERS Extraction Schemes**

PAH	3% <sup>a</sup>		8%		11%	
	E3	E4	E3	E4	E3	E4
naphthalene	49.1 $\pm$ 5.9	52.6 $\pm$ 4.0	59.6 $\pm$ 2.5	62.9 $\pm$ 2.2	46.1 $\pm$ 2.7	48.6 $\pm$ 7.7
2-methylnaphthalene	24.5 $\pm$ 3.9	26.7 $\pm$ 2.9	38.5 $\pm$ 0.6	38.6 $\pm$ 2.3	15.8 $\pm$ 0.9	17.2 $\pm$ 3.4
1-methylnaphthalene	11.5 $\pm$ 1.3	13.0 $\pm$ 1.6	18.0 $\pm$ 1.5	17.6 $\pm$ 1.6	8.0 $\pm$ 0.6	9.0 $\pm$ 1.6
phenanthrene	12.0 $\pm$ 1.3	13.8 $\pm$ 0.5	11.3 $\pm$ 0.2	12.6 $\pm$ 0.7	11.2 $\pm$ 0.6	11.2 $\pm$ 0.8
anthracene	BDL <sup>b</sup>	BDL	ND <sup>c</sup>	ND	ND	ND
fluoranthene	7.5 $\pm$ 1.1	9.3 $\pm$ 0.4	8.9 $\pm$ 0.6	9.1 $\pm$ 0.1	7.6 $\pm$ 1.7	8.4 $\pm$ 1.7
pyrene	24.7 $\pm$ 6.7	28.5 $\pm$ 1.5	22.9 $\pm$ 2.7	28.9 $\pm$ 1.3	26.4 $\pm$ 6.6	29.4 $\pm$ 6.8
benzo[ <i>g,h,i</i> ]perylene	7.2 $\pm$ 3.9	6.0 $\pm$ 0.6	5.4 $\pm$ 0.9	4.3 $\pm$ 0.9	6.9 $\pm$ 1.3	7.2 $\pm$ 1.3
$\Sigma$ PAHs <sup>d</sup> (ng/g wet weight)	137 $\pm$ 11	150 $\pm$ 6	165 $\pm$ 4	174 $\pm$ 4	122 $\pm$ 8	131 $\pm$ 11

<sup>a</sup> Fat content. <sup>b</sup> BDL: compound met all quantitation criteria, but S/N < 3. <sup>c</sup> ND: the compound was not detected; PAHs absent from the table were not detected. <sup>d</sup>  $\Sigma$ PAHs represents the summation of PAHs with levels above detection limits. PAHs absent from the table were not detected in salmon samples.

platform implementing a three-component acetone, ethyl acetate and isooctane extraction solvent in a 2:2:1 (v/v/v) ratio coupled to dispersive SPE sample cleanup and GC-MS is a fast, selective, efficient and precise method for the determination of PAHs in high-fat smoked fish products. The modified QuEChERS methods described show good potential for use in monitoring levels of PAHs in lipid-rich fish.

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## ABBREVIATIONS USED

AOAC, Association of Official Analytical Chemists; ASE, accelerated solvent extraction; EN, European Committee for Standardization; GC-MS, gas chromatography-mass spectrometry; PAH, polycyclic aromatic hydrocarbon; PSA, primary, secondary amine; QuEChERS, quick, easy, cheap, effective, rugged and safe; SIM, selective ion monitoring; SPE, solid-phase extraction; TIC, total ion current

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